

Available online at www.sciencedirect.com

Tetrahedron Letters 45 (2004) 4277–4280

Tetrahedron Letters

Stereoselective synthesis of novel conformationally restricted β - and γ -amino acids

Frieder Gnad, Marko Poleschak and Oliver Reiser*

[I](mail to: oliver.reiser@chemie.uni-regensburg.de
)nstitut für Organische Chemie, Universität Regensburg, Universitätsstr. 31, Regensburg D-93040, Germany

Received 13 February 2004; revised 5 April 2004; accepted 5 April 2004

Dedicated to Professor Armin de Meijere on the occasion of his 65th birthday

Abstract—Novel conformationally restricted β - and γ -amino acids containing a cyclopropane ring could be stereoselectively synthesized from readily available 5-methoxyindole and pyridine by copper(I)-catalyzed cyclopropanation with methyl diazoacetate followed by subsequent oxidative cleavage of the resulting adducts. 2004 Elsevier Ltd. All rights reserved.

The introduction of conformationally restricted amino acids into peptides is an important tool in the elucidation of the biologically active conformation of proteins.¹ Proline, α -aminoisobutyric acid $(AIB)^2$ and α -aminocyclopropane carboxylic acid³ $(\alpha$ -ACC) are three prominent examples for such amino acids, which have been used extensively in the design of peptidomimetics directed towards the development of new drugs (Fig. 1).

Recently, B-aminocyclopropanecarboxylic acids⁴ (β-ACCs), especially the *cis*-β-ACC 3b,⁵ have been recognized as uniquely constrained building blocks in the synthesis of peptidic ligands, for example, with high affinity towards the neuropeptide Y receptors 6 and in the synthesis of helical α, β -foldamers.⁷ Both, *cis*- and trans- β -ACCs 3 can be conveniently synthesized^{5a} from N-Boc-pyrrole 1 via cyclopropanation with methyl diazoacetate followed by ozonolysis of the bicyclic adduct 2 (Fig. 2) and subsequently incorporated into

Figure 1. Conformationally restricted α -amino acids.

0040-4039/\$ - see front matter \odot 2004 Elsevier Ltd. All rights reserved. doi:10.1016/j.tetlet.2004.04.013

peptides using an in situ Boc^{5b} or Alloc⁸ coupling approach.

In extension of this strategy we report here the cyclopropanation of indoles and dihydropyridines that leads to novel β - and γ -amino acids, being conformationally restricted by the incorporation of a cyclopropane moiety into the backbone.

Following the pioneering work of Welstead et al.⁹ and of Wenkert et al.,10 cyclopropanation of N-Boc protected indoles 4 with methyl diazoacetate catalyzed by copper(I)triflate, being generated in situ from copper(II)triflate and phenylhydrazine, led to the expected $[2+1]$ cycloadducts of the double bond in the five-membered ring (Scheme 1).

Both indoles 4 gave the cyclopropanated adducts in a clean reaction. Initially only 1.4 equiv of methyl diazoacetate were used, which resulted in an incomplete conversion of $4a$, but nevertheless, high yield of $\bar{5}a/6a$ after recovery of the starting material (90%). When a

Keywords: Amino acids; Cyclopropanes; Peptidomimetics; Catalysis; Pyridines; Indoles.

^{*} Corresponding author. Tel.: +49-941-9434631; fax: +49-941-94341 21; e-mail: [oliver.reiser@chemie.uni-regensburg.de](mail to: oliver.reiser@chemie.uni-regensburg.de
) Figure 2. Synthesis of β -ACCs 3.

Scheme 1. Cyclopropanation of indoles 4. (a) N_2CHCO_2Me (4a: 1.4 equiv, $4b$: 7.3 equiv), $Cu(OTf)_2$ (0.02 equiv), PhNHNH₂, (0.02 equiv) , $CH₂Cl₂$.

large excess (7.3 equiv) of methyl diazoacetate was employed in the cyclopropanation of 4b, a nearly quantitative yield of 5b/6b was obtained. However, in contrast to the cyclopropanation of 1, for which only the exoadduct 2 was observed, with 4 both, the exo-adducts 5 as well as the *endo*-adducts **6** were obtained $(2-3:1)$. In both cases, the diastereomers could be readily separated by chromatography on silica and individually investigated further.

Oxidative degradation of either 5a or 6a was not successful under various conditions tested. However, the more electron rich aromatic moiety in the derivatives 5b and 6b could be both cleaved by ozone, however, with a very different outcome depending on the stereochemistry of the adducts (Scheme 2). The exo-stereoisomer 5b was converted to the expected pyrrolidone 7, in which complete oxidative cleavage of the arene moiety has been taken place. In contrast, the endo-stereoisomer 6b

Scheme 2. Ozonolysis of cyclopropanated indoles 5b and 6b. (a) (i) O_3 , MeOH, -78 °C ; (ii) Me₂S, $-78 \text{ °C} \rightarrow$ room temp, $5b \rightarrow 7$: 56%, $6b \rightarrow 8$: 54%.

Scheme 3. Ring opening of 7. (a) Morpholine (1 or 2 equiv), CH_2Cl_2 , 0° C \rightarrow room temp, 35%; (b) MeOH (aq), NH₄Cl, 45 °C, 3h, 63%.

gave in reasonable yield the partial oxidized alkene 8 as the sole product.

Following the precedent set for isatins, $\frac{11}{11}$ ring opening of 7 could be effected either with a secondary amine or with hydroxide (Scheme 3). With morpholine, two-fold addition giving rise to 9 occurred, which was surprisingly stable in spite of its hemi aminal functionality. Alternatively, the ketoacid 10^{12} was cleanly formed with ammonium chloride in methanol, representing a novel γ -amino acid that should prove to be interesting as a complimentary building block to the $trans- β -ACC 3a$.

Starting from dihydropyridine 11, which can be conveniently synthesized directly from pyridine following the protocol reported in the literature,¹³ novel β - and γ amino acids containing a cyclopropane ring can also be readily obtained (Scheme 4). Cyclopropanation¹⁴ of 11 with methyl diazoacetate as described before for indoles 4 gave rise to both, exo- and endo (ratio 8:3) monocyclopropanated adducts 12 and 13 (55% overall yield), respectively, which were separated by chromatography on silica. Ozonolysis of the individual isomers followed by reductive work up cleanly gave rise to the aldehydes 14 and 15, respectively, which should prove to be valuable, trifunctional building blocks not only in light of the synthesis of unnatural amino acids described here. Nevertheless, oxidation using sodium chlorite/hydrogen peroxide according to the Dalcanale protocol¹⁵ proceeded in an almost quantitative way to the new cis - γ amino acids 16 and 17 , complementing the cis- β -ACC 3b.

Alternatively, the *trans*- β -ACC 19 can be obtained by saponification of 12 followed by ozonolysis and reductive work up (Scheme 5). It should be noted that the direct saponification of 14 with base towards 19 results only in decomposition of the starting material.

The aldehyde group in 14 can be functionalized by standard transformations, which can be applied to the synthesis of β -ACCs with α -amino acid side chain functionality.16 Reductive amination with concomitant deprotection of the N-formyl group leads to the deriv-

Scheme 4. Synthesis of γ -amino acids containing a cyclopropane ring from pyridine. (a) Ref. 8; (b) N_2CHCO_2R (1.4 equiv), $Cu(OTf)_2$ (0.05 equiv), PhNHNH₂, CH₂Cl₂, room temp, 12 (40%), 13 (15%); (c) O₃, DMS (5 equiv), CH₂Cl₂, $-78 \text{ °C} \rightarrow$ room temp, **14** (91%), **15** (91%); (d) (i) NaClO₂, H₂O₂ (1.1 equiv), CH₃CN, 0° C \rightarrow room temp; (ii) 2diethylaminoethylamine (DEAEA, 2.2 equiv), CH₃CN, room temp, 10–12 h, yield (two steps): 16 (84%), 17 (72%).

ative 20 , representing a *trans*- β -ACC derivative with a lysine side chain (Scheme 6).

In conclusion, we have demonstrated that pyridine and 5-methoxyindole can be readily transformed into a variety of new β - and γ -amino acids containing a cyclopropane moiety, which further extends the scope of this class of conformationally restricted amino acids as building blocks for peptidomimetics.

19

Scheme 5. Synthesis of trifunctional trans- β -ACC 19. (a) NaOH (1.25 equiv), MeOH/H₂O, room temp, 89%; (b) O_3 , DMS (5 equiv), CH_2Cl_2 , $-78 \text{°C} \rightarrow$ room temp, 93%.

Scheme 6. Synthesis of the β -ACC derivative 20 with lysine side chain functionality. (a) benzylamine $(2.2 \text{equiv}), \text{NaBH}(\text{OAc})$ ₃ $(2.5 \text{equiv}),$ molecular sieves (4 Å), THF, $0^{\circ}C \rightarrow$ room temp 12 h, 47%.

Acknowledgements

This work was supported by the Deutsche Forschungsgemeinschaft (RE 948-4/2 and the Graduiertenkolleg Medizinische Chemie GRK 760), the Fonds der Chemischen Industrie, and through generous gifts of chemicals by Degussa AG.

References and notes

- 1. Bailey, P. D. An Introduction to Peptide Chemistry; Wiley: New York, 1990.
- 2. (a) Toniolo, C.; Bonora, G. M.; Bavoso, A.; Benedetti, E.; di Blasio, B.; Pavone, V.; Pedone, C. Biopolymers 1983, 22, 205–215; (b) Benedetti, E.; di Blasio, B.; Pavone, V.; Pedone, C.; Santini, A.; Crisma, M.; Valle, G.; Toniolo, C. Biopolymers 1989, 28, 175–184; (c) Basu, G.; Bagchi, K.; Kuki, A. Biopolymers 1991, 31, 1763–1774; (d) Karle, I. L.; Balaram, P. Biochemistry 1990, 29, 6747–6756; (e) Karle, I. L. Biopolymers 1996, 40, 157–180; (f) Marshall, G. R.; Hodgkin, E. E.; Langs, D. A.; Smith, G. D.; Zabrocki, J.; Leplawy, M. T. Proc. Natl. Acad. Sci. 1990, 87, 487–491.
- 3. (a) Tsang, J. W.; Schmeid, M.; Nyfelter, R.; Goodman, M. J. J. Med. Chem. 1984, 27, 1663; (b) Mapelli, C.; Stammer, C. H.; Lok, S.; Mierke, D. F.; Goodman, M. Int. J. Pept. Protein Res. 1988, 32, 484; (c) Zhu, Y. F.; Yamazaki, T.; Tsang, J. W.; Lok, S.; Goodman, M. J. Org. Chem. 1992, 57, 1074; (d) Ner, S. K.; Suckling, C. J.; Bell, A. R.; Wrigglesworth, R. J. Chem. Soc., Chem. Commun. 1987, 23, 480–482; (e) Kimura, H.; Stammer, C. H.; Shimohigashi, Y.; Cui, R. L.; Stewart, J. J. Biochem. Biophys. Res. Commun. 1983, 115, 112; (f) Shimohigashi, Y.; Stammer, C. H.; Costa, T.; Voigtlander, V. P. F. Int. J. Pept. Protein Res. 1983, 22, 489; (g) Shimohigashi, Y.; Costa, T.; Nitz, T. J.; Chen, H. C.; Stammer, C. H. Biochem. Biophys. Res. Commun. 1984, 121, 966; (h) Mapelli, C.; Kimura, H.; Stammer, C. H. Int. J. Pept. Protein Res. 1986, 28, 347; (i) Ahmad, S.; Phillips, R. S.; Stammer, C. H. J. Med. Chem. 1992, 35, 1410; (j) Burgess, K.; Ho, K.-K.; Pettitt, B. M. J. Am. Chem. Soc. 1995, 117, 54–65; (k) Burgess, K.; Ho, K.-K. J. Am. Chem. Soc. 1994, 116, 799–800; (l) Malin, D. H.; Payza, K.; Lake, J. R.; Corriere, L. S.; Benson, T. M.; Smith, D. A.; Baugher, R. K.; Ho, K.-K.; Burgess, K. Peptides 1993, 14, 47; (m) Malin, D. H.; Lake, J. R.; Ho, K.-K.; Corriere, L. S.; Garber, T. M.; Waller, M.; Benson, T.; Smith, D. A.; Luu, T.-A.; Burgess, K. Peptides 1993, 14, 731; (n) Moye-Sherman, D.; Jin, S.; Li, S.; Welch, M. B.; Reibenspies, J.; Burgess, K. Chem. Eur. J. 1999, 5, 2730–2739.
- 4. Review: Gnad, F.; Reiser, O. Chem. Rev. 2003, 103, 1603– 1624; (a) Representative examples: Paulini, K.; Reissig, H.-U. Liebigs Ann. Chem. 1994, 549–554; (b) Beck-Sickinger, A. G.; Hoffmann, E.; Paulini, K.; Reissig,

H.-U.; Willim, K.-D.; Wieland, H. A.; Jung, G. Biochem. Soc. Trans. 1994, 22, 145-149; (c) Hibbs, D. E.; Hursthouse, M. B.; Jones, I. G.; Jones, W.; Malik, K. M. A.; North, M. Tetrahedron 1997, 53, 17417–17424; (d) North, M. J. Pept. Sci. 2000, 6, 301–313.

- 5. (a) Beumer, R.; Bubert, C.; Cabrele, C.; Vielhauer, O.; Pietzsch, M.; Reiser, O. J. Org. Chem. 2000, 65, 8960– 8969; (b) Voigt, J.; Noltemeyer, M.; Reiser, O. Synlett 1997, 202–204; (c) Bubert, C.; Cabrele, C.; Reiser, O. Synlett 1997, 827–829.
- 6. Koglin, N.; Zorn, C.; Beumer, R.; Cabrele, C.; Bubert, C.; Sewald, N.; Reiser, O.; Beck-Sickinger, A. G. Angew. Chem., Int. Ed. Engl. 2003, 42, 202–205.
- 7. De Pol, S.; Zorn, C.; Klein, C.; Zerbe, O.; Reiser, O. Angew. Chem., Int. Ed. Engl. 2004, 43, 511–514.
- 8. Zorn, C.; Gnad, F.; Salmen, S.; Herpin, T.; Reiser, O. Tetrahedron Lett. 2001, 42, 7049–7053.
- 9. Cyclopropanation of N-carbomethoxyindole with ethyl diazoacetate using CuCN has been reported: Welstead, W. J., Jr.; Stauffer, H. F., Jr.; Sancilio, L. F. J. Med. Chem. 1974, 17, 544–547.
- 10. Cyclopropanation of Z-protected indoles with ethyl diazoacetate using copper bonze has been reported: Wenkert, E.; Alonso, M. E.; Gottlieb, H. E.; Sanchez, E. L.; Pellicciari, R.; Cogolli, P. J. Org. Chem. 1977, 42, 3945– 3949; for an alternative approach towards cyclopropanated indoles see: Li, C.; Vasella, A. Helv. Chim. Acta 1993, 76, 197–210.
- 11. (a) Franke, A. Liebigs Ann. Chem. 1982, 23, 794–804; (b) Black, D. St. C.; Moss, G. I. Aust. J. Chem. 1987, 40, 129– 142; (c) Meyer, F. I. Chem. Ber. 1966, 99, 3060–3062; (d) Johnsen, B. A.; Undheim, K. Acta Chem. Scand. Ser. B 1984, 38, 109–112;

(e) Catto, A.; Cappelletti, R.; Leonardi, A.; Tajana, A.; Maggi, F.; Nardi, D.; Taddei, F. Farmaco Ed. Sci. 1983, 38, 45–56; (f) Nardi, D.; Tajana, A.; Portioli, F.; Bonola, G. Farmaco Ed. Sci. 1982, 37, 815–823; (g) Egli, R.; Richter, C. Helv. Chim. Acta 1957, 499–501; (h) Tomchin, A. B.; Marysheva, V. V. Zh. Org. Khim. 1993, 29, 444–446;

(i) Tomchin, A. B. Zh. Org. Khim. 1990, 26, 860–873; (j) Tomchin, A. B. Zh. Org. Khim. 1987, 23, 1305–1312.

- 12. All new compounds were characterized by IR, NMR and MS and gave sufficient elemental analyses or high resolution mass spectra. Selected ¹H NMR data. Compound 10: ¹H NMR (CDCl₃, signal doubling due to rotamers) δ 1.54 $(9H, s, -CO_2C(CH_3)$ ₃, 1.86–1.94 (m, 1H, cyclopropyl-H), 2.30–2.42 (m, 1H, cyclopropyl-H), 3.71 and 3.72 (s, 3H, $-CO₂CH₃$), 3.95–4.02 (m, 1H, cyclopropyl-H). Compound 16: ¹H NMR (CDCl₃) δ 1.44 (s, 9H, -CO₂C(CH₃)₃), 1.57– 1.61 (m, 1H, cyclopropyl-H), 1.77–1.83 (m, 1H, cyclopropyl-H), 2.39–2.56 (m, 2H, CH2), 3.02–3.07 (m, 1H, cyclopropyl-H), 3.68 (s, 3H, OCH₃). Compound 17: ¹H NMR (CDCl₃) δ 1.44 (s, 9H, $-CO_2C(CH_3)_{3}$), 1.77–1.85 (m, 1H, cyclopropyl-H), 1.98–2.05 (m, 1H, cyclopropyl-H), 2.71 (d, J 7.6 Hz, 2H, CH₂), 3.13 (dd, J 7.4, 7.5 Hz, 1H, cycopropyl-H), 3.63 (s, 3H, OCH₃). Compound 19: ¹H, NMR (CDCl₃) δ 1.53 (s, 9H, –CO₂C(CH₃)₃), 1.76 (dd, J 4.3, 5.5 Hz, 1H, cyclopropyl-H1), 1.89–2.19 (m, 2H, CH2), 2.95–3.04 (m, 1H, cyclopropyl-H), 3.43 (dd, J 4.1, 7.3 Hz, 1H, cyclopropyl-H), 9.12 (s, 1H, N–CHO), 9.69 (s, 1H, CHO). Compound 20: ¹H NMR (CDCl₃) δ 1.41 (s, 9H, $-CO_2C(CH_3)$ ₃), 1.62–1.79 (m, 3H, CH₂ and cyclopropyl-H), $1.87-1.98$ (m, $2H$, CH₂) $2.78-2.86$ (m, 1H, cyclopropyl-H), 3.31–3.39 (m, 1H, cyclopropyl-H), 3.63 (s, 1H, OCH3), 3.89 (br s, 2H, CH2Ph), 7.1–7.32 (m, 5H, phenyl-H).
- 13. (a) Fowler, F. W. J. Org. Chem. 1972, 37, 1321–1323; (b) Sundberg; Bloom J. Org. Chem. 1981, 46, 4836–4842.
- 14. Cyclopropanation of substituted 1,4-dihydropyridines with ethyl diazoacetate: Wenkert, E.; Broquet, C. Synth. Commun. 1979, 9, 689–695; (a) Simmons–Smith cyclopropanation of unsubstituted 1,4-dihydropyridines: Bubert, C.; Voigt, J.; Biasetton, S.; Reiser, O. Synlett 1994, 8, 675–677.
- 15. Review: Raach, A.; Reiser, O. J. Prakt. Chem. 2000, 342, 605–608.
- 16. Cf. Beumer, R.; Reiser, O. Tetrahedron 2001, 57, 6497– 6503.